

A correlation between two different species of fish embryos used in a freshwater qualitative pollution test

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Abstract

*Pollutants of freshwater sources, through their eco-risk potential can affect the human and animal health, numerous risk-assays being imagined and applied in the last decade. In this respect our aim was to make, an initial acute eco-risk study comparing the zebrafish embryos with an of interest autochthon species, namely pikeperch embryos, to see if this species is suitable and can be introduced as reliable testing tool in temperate region's water eco-pollution studies. The ISO 15088:2007 test method, adapted by our collective to pikeperch embryos stage development was used. Accordingly, 100 zebrafish embryos (*Danio rerio*) and 20 as control, in parallel with 100 pikeperch embryos (*Sander lucioperca*) and 20 control were kept in water samples provided from a swine farm surroundings, diluted in ten different concentrations: 1X, 2X, 3X, 4X, 6X, 8X, 12X, 16X, 24X, 32X. Any behaviour changes and mortality was recorded and statistically analysed according to Kolmogorov-Smirnov normality test, where P value was significant when $P < 0.05$. Though pikeperch embryo use for testing are not a pattern yet, as zebrafish embryos, results revealed that the indigene species embryos can be successfully used in studies applicable in ecotoxicology or other biomedical domains, with comparable results to the classical one.*

Keywords: acute embryo-toxicity, pikeperch, zebrafish, risk-assay.

1. Introduction

In eco-toxicology and biomedical domains zebrafish adults and embryos are successfully used, in a large palette of diagnostic and experimental models, in order to test various substances, from environmental pollutants to the active medicinal substances. From these, the acute embryo tests are helpful in providing an easily available, low-cost option, which can give enough accurate results in a limited amount of time [1-15]. Toxicity effects of diverse components in freshwaters was highlighted by the changes in anatomical and ethologic development of the zebrafish embryonic and larval stages, their eggs being increasingly used in current pollution hazard and risk assessments, because of their qualities: transparency, lack of adhesion, reduced diameter, that give them a great availability and reproducibility [16-19].

Based on the fact that, acute toxicity tests on fish embryos can be successfully used as indicators of the river's pollution, including with animal effluents, and our observations made in different fish species embryos, that revealed useful similarities between zebrafish and another species, more common and of great interest for the temperate climate, namely the

pikeperch embryos (*Sander lucioperca*), we have tried to extrapolate this bio-methodology from zebrafish to pikeperch embryos.

In addition, our study was also justified due to the large producing pikeperch countries (e.g. Czech Republic, Denmark, Hungary, Romania, Tunisia, Ukraine) [20], great scientific interests in this species, this being another reason why a comparative study it should have been made, in the aim to develop a risk assessment tool, helpful for ecotoxicology, the fish industry and/or public health domains.

2. Material and methods

Samples collection and methodology

Water samples for this test were obtained from Ciacova, Timiș County, Romania. The place was chosen due to the river Timiș placement, in the near vicinity of the swine farm.

Previously to this test we have measured after the known methodology the main physicochemical parameters, including nitrate; chloride and phosphate levels (with values within the European legislation limits) [21] (Table 1).

Table 1. Main indicators and parameters for the sampling site

Parameters		Values
Location GPS coordinates	Long.	45°51'65"
	Lat.	21°1'49"
Water temperature (°C)		9.4
Ambient temperature (°C)		8
Atmospheric pressure (hPa)		1012
Weather condition		Cloudy
Relative humidity (%)		63
Conductivity (μS)		730.70
Salinity (mg/L)		351.0 (0.4 ‰)
pH		7.7
Dissolved oxygen (mg/L)		9.45
Chloride (ppm)		120.0
Nitrate (mg/dm ³)		1.797
Phosphate (mg/dm ³)		0.0175

In this initial assay there were used 100 zebrafish embryos (*Danio rerio* Hamilton-Buchanan) and 20 controls in parallel with 100 pikeperch embryos (*Sander lucioperca* L.) and 20 controls.

The common test method used, was ISO 15088:2007: Water quality - Determination of the acute toxicity of waste water to zebrafish eggs (*Danio rerio*), a rapid risk-assay, this standard being reviewed and confirmed in 2011, adapted by us for the pikeperch, according to their species stage development. According to its description, this International Standard specifies a method for the determination of degrees of dilution or of concentrations as a measure of the acute toxic effect of waste water to fish eggs within 48 h. This International Standard is also applicable to treated municipal waste water and industrial effluents [22].

In both tests, using zebrafish and pikeperch embryos, there were used ten dilutions respectively: 1X, 2X, 3X, 4X, 6X, 8X, 12X, 16X, 24X, 32X. As negative control, it was used system water. Fish embryos were placed in 24-well plates (one embryo in one well) and incubated. Unlike zebrafish embryos, incubated at 26°C, pikeperch embryos were incubated at 18°C, until there were obtained 80°C cumulative degrees until fish hatching. All behavioural and developmental changes were monitored for the entire test duration and the standard critical endpoints were noted for each sample.

3. Statistics

Obtained data was analysed for normal distribution with Kolmogorov - Smirnov (K-S) normality test using Minitab Statistical Software version 16 (Minitab Inc., UK). The non-parametric test K-S test compare one or two samples with a reference probability distribution being one of the most useful for comparing two samples, as it is sensitive to differences in both location and shape of the empirical cumulative distribution functions of the two samples, the results being considered significant when $P < 0.05$.

4. Results and Discussions

In this initial assay, from day 0 post-fecundation to the standard defined endpoints: coagulated eggs, lack of heartbeat, developmental tail and larval disorders, there were observed and registered comparatively for both fish species and for all tested samples (Fig 1). Our observations revealed a great similitude between pikeperch and zebrafish as evolution of the developmental changes including here also the eggs behaviour to the tested concentrations.

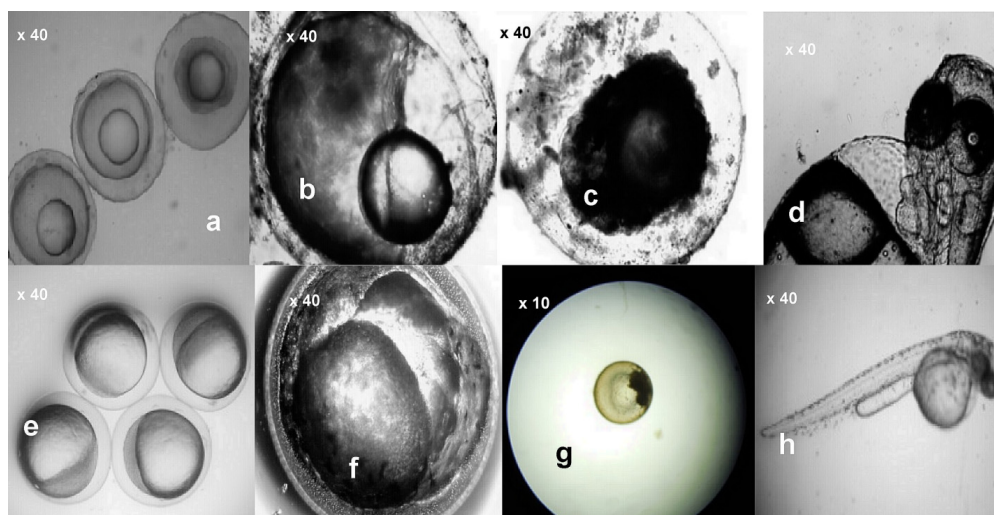


Figure 1. pikeperch vs. zebrafish: (a)/(e) - eggs on day 0 post-fecundation; (b)/(f) - undeveloped embryos; (c)/(g) coagulated eggs, (d)/(h) larvae neo-formation bodies.

Additionally to the endpoints, in both fish embryo species samples, there were observed similar mortality rates for the great concentrations: 16X, 24X and 32X (Fig 2). According to K-S normality statistical test, P value obtained was significant ($P = 0.047$) for zebrafish embryo mortality (Fig 3), while pikeperch embryo mortality had no statistical significance in our case ($P = 0.372$).

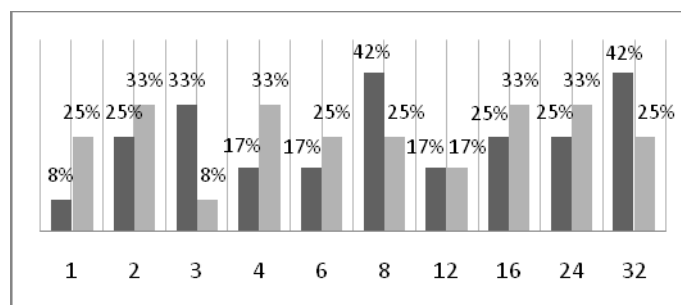


Figure 2. Mortality percentage of Pikeperch (black) and Zebrafish (gray) embryos

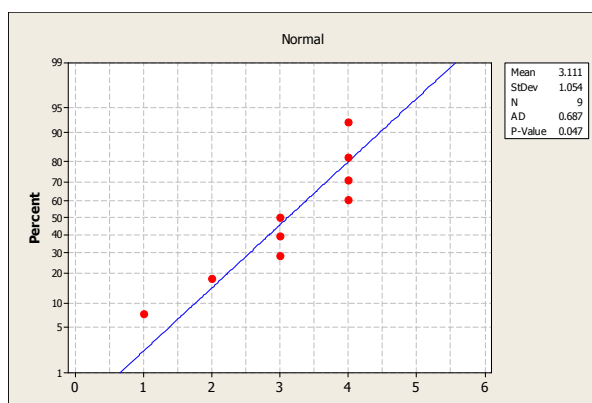


Figure 3. Statistical interpretation of zebrafish embryo sample

In the last decade, studies on zebrafish diversified much, bringing new information, especially on the fields of environmental, aquatic, and reproductive (biomarkers) toxicology [23- 25]. Among these, embryo-tests showed that they are sensitive enough to detect pollutants in wastewaters both in acute and long-term studies, more and more researchers putting the problem of using the embryo toxicity test (FET) (as alternative to the fish acute toxicity test), being a reliable tool to assess the embryo toxicity potential, affirmation that we fully agree [26-28].

In this aim, also a notable advantage of this testing methodology, is that according to the European welfare regulations, zebrafish embryo testing is categorized as an *in vitro* testing, thus not being subject to the animal protection legislation [29, 30], situation that, in our eyes, can be easily extrapolated to the pikeperch embryos and further develop the studies on this species.

Knowing that zebrafish have a very similar development to mammals becoming an accessible model in research, we supposed correctly that pikeperch embryos also can have such feature more suitable to temperate conditions, being a suitable and possible replacer for the zebrafish embryos. Comparative zebrafish vs. pikeperch studies are at the beginning, only a few communications being available in the literature until now.

In pikeperch it was yet established using same methodology as for zebrafish, that nitrogenous compounds have an impact in the development of fishes, the early life-stage forms being most susceptible [31, 32]. From other previous studies made, it was highlighted until now, that pikeperch is rather susceptible to nitrite in comparison with zebrafish [33]. Also bio substances accumulation in pikeperch tissues appreciating their impact in case of consumption by humans was established by some authors using zebrafish methodology [34-36].

5. Conclusions

Although pikeperch embryos use are not yet a pattern, like zebrafish embryos with large applications, this species embryos have been shown to be successfully used in comparative studies. In our opinion pikeperch embryos can be considered fully experimental model organisms and a risk assessment tool, helpful for ecotoxicology, the fish industry and/or public health domains.

In our opinion the main advantages of implementing a 48 hours acute toxicity risk-assay for pikeperch embryos could be: high numbers of embryos that can be used with low costs, efficiency results revealed in a short period of time, fast and precise response to the freshwater and eco-environments pollutants, medicinal active substances or heavy metals.

This eco-toxicity method could be used as an alternative assay in the fish industry for the pikeperch breed in the recirculating water systems.

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References

- (1) T. BRAUNBECK, M. BÖTTCHER, H. HOLLERT, T. KOSMEHL, E. LAMMER, E. LEIST, M. RUDOLF, N. SEITZ, Towards an alternative for the acute fish LC₅₀ test in chemical assessment: the fish embryo toxicity test goes multi-species - an update. *ALTEX*, 22(2), 87–102, (2005).
- (2) F. BUSQUET, R. NAGEL, F. VON LANDENBERG, S.O. MUELLER, N. HUEBLER, T.H. BROSCARD, Development of a new screening assay to identify pro teratogenic substances using zebrafish *Danio rerio* embryo combined with an exogenous mammalian metabolic activation system (mDarT). *Toxicol Sci*, 104(1), 177-188, (2008).
- (3) C. CHAKRABORTY, C.H. HSU, Z.H. WEN, C.S. LIN, G. AGORAMOORTHY, Zebrafish: a complete animal model for in vivo drug discovery and development. *Curr Drug Metab*, 10(2), 116-124, (2009).
- (4) R.M. EMBRY, S.E. BELANGER, T.A. BRAUNBECK, M. GALAY-BURGOS, M. HALDER, D.E. HINTON, M.A. LÉONARD, A. LILICRAP, T. NORBERG-KING, G. WHALE, The fish embryo toxicity test as an animal alternative method in hazard and risk assessment and scientific research. *Aquat Toxicol*, 97(2), 79-87, (2010).
- (5) A.J. HILL, H. TERAOKA, W. HEIDEMAN, R.E. PETERSON, Zebrafish as a model vertebrate for investigating chemical toxicity. *Toxicol Sci*, 86(1), 6-19, (2005).
- (6) E. LAMMER, H.G. KAMP, V. HISGEN, M. KOCH, D. REINHARD, E.R. SALINAS, K. WENDLER, S. ZOK, T. BRAUNBECK, Development of a flow-through system for the fish embryo toxicity test (FET) with the zebrafish (*Danio rerio*). *Toxicol In Vitro*, 23(7), 1436-1442, (2009).
- (7) E. LOUCKS, M.J. CARVAN, Strain-dependent effects of developmental ethanol exposure in zebrafish. *Neurotoxicol Teratol*, 26(6) 745-755, (2004).
- (8) P. MCGRATH, C.Q. LI, Zebrafish: a predictive model for assessing drug-induced toxicity. *Drug Discov Today*, 13(9-10), 394-401, (2008).
- (9) K. SCHIRMER, K. TANNEBERGER, N.I. KRAMER, D. VOLKER, S. SCHOLZ, C. HAFNER, L.E. LEE, N.C. BOLS, J.L. HERMENS, Developing a list of reference chemicals for testing alternatives to whole fish toxicity tests. *Aquat Toxicol*, 90(2), 128-137, (2008).
- (10) Z.G. LU, M.H. LI, J.S. WANG, D.D. WEI, Q.W. LIU, L.Y. KONG, Developmental toxicity and neurotoxicity of two matrine-type alkaloids, matrine and sophocarpine, in zebrafish (*Danio rerio*) embryos/larvae. *Reprod Toxicol*, 47, 33-41, (2014).
- (11) S.H. SEOK, M.W. BAEK, H.Y. LEE, D.J. KIM, Y.R. NA, K.J. NOH, S.H. PARK, H.K. LEE, B.H. LEE, J.H. PARK, In vivo alternative testing with zebrafish in ecotoxicology. *J Vet Sci*, 9(4), 351-357, (2008).
- (12) C. TON, Y. LIN, C. WILLETT, Zebrafish as a model for developmental neurotoxicity testing. *Birth Defects Res A Clin Mol Teratol*, 76(7), 553–567, (2006).
- (13) L. YANG, N.Y. HO, R. ALSHUT, J. LEGRADI, C. WEISS, M. REISCHL, R. MIKUT, U. LIEBEL, F. MÜLLER, U. STRÄHLE, Zebrafish embryos as models for embryotoxic and teratological effects of chemicals. *Reprod Toxicol*, 28(2), 245–253, (2009).
- (14) S. ALI, D.L. CHAMPAGNE, H.P. SPAINK, & RICHARDSON, M.K. Zebrafish embryos and larvae: a new generation of disease models and drug screens. *Birth Defects Res C Embryo Today Rev*, 93(2), 115-133, (2011).
- (15) K.C. BRANNEN, J.M. PANZICA-KELLY, T.L. DANBERRY, K.A. AUGUSTINE-RAUCH, Development of a zebrafish embryo teratogenicity assay and quantitative prediction model. *Birth Defects Res B Dev Reprod Toxicol*, 89(1), 66–77, (2010).
- (16) M. LANGE, W. GEBAUER, J. MARKL, R. NAGEL, Comparison of testing acute toxicity on embryo of zebrafish (*Brachydanio rerio*), and RGT-2 cytotoxicity as possible alternatives to the acute fish test. *Chemosphere*, 30(11), 2087-2102, (1995).

A correlation between two different species of fish embryos
used in a freshwater qualitative pollution test

- (17) S. SCHOLZ, S. FISCHER, U. GÜNDEL, E. KÜSTER, T. LUCKENBACH, D. VOELKER, The zebrafish embryo model in environmental risk assessment--applications beyond acute toxicity testing. *Environ Sci Poll Res Int*, 15(5), 394-404, (2008).
- (18) I.W. SELDERSLAGHS, A.R. VAN ROMPAY, W. DE COEN, H.E. WITTERS, Development of a screening assay to identify teratogenic and embryotoxic chemicals using the zebrafish embryo. *Reprod Toxicol*, 28(3) 308–320, (2009).
- (19) S. WEIGT, N. HUEBLER, T. BRAUNBECK, F. VON LANDENBERG, T.H. BROSCARD, Zebrafish teratogenicity test with metabolic activation (mDarT): effects of phase I activation of acetaminophen on zebrafish *Danio rerio* embryos. *Toxicology*, 275(1-3), 36–49, (2010).
- (20) *FAO Fisheries & Aquaculture Sander lucioperca*, Available at: http://www.fao.org/fishery/culturedspecies/Sander_lucioperca/en
- (21) C.L. MOȘNEANG, V.L. ORDODI, R.T. CRISTINA, An analysis of water samples surrounding swine farms in Timiș County – A practical guide. *Medicamentul Veterinar / Veterinary Drug*, 7(2), 56-85, (2013).
- (22) ISO 15088:2007: Water quality - Determination of the acute toxicity of waste water to zebrafish eggs (*Danio rerio*)
- (23) K. BAKOS, R. KOVACS, A. STASZNY, D. KANAINÉ SIPOS, B. URBANYI, F. MULLER, Z. CSENKI, B. KOVACS, Developmental toxicity and estrogenic potency of zearalenone in zebrafish (*Danio rerio*). *Aquat Toxicol*, 136-137, 13-21 (2013).
- (24) A.V. HALLARE, M. SCHIRLING, T. LUCKENBACH, H.R. KÖHLER, R. TRIEBSKORN, Combined effects of temperature and cadmium on developmental parameters and biomarker responses in zebrafish (*Danio rerio*) embryos, *J Therm Biol*, 30, 7–17, (2005).
- (25) W.H. LI, P.C. CHAN, K.M. CHAN, Metal uptake in zebrafish embryo-larvae exposed to metal-contaminated sediments, *Mar Environ Res*, 58, 829–832, (2004).
- (26) K. HENN, T. BRAUNBECK, Dechorionation as a tool to improve the fish embryo toxicity test (FET) with the zebrafish (*Danio rerio*). *Comp Biochem Physiol C: Pharmacol Toxicol Endocrinol*, 153, 91-98, (2011).
- (27) E. LAMMER, G.J. CARR, K. WENDLER, J.M. RAWLINGS, S.E. BELANGER, T. BRAUNBECK, Is the fish embryo toxicity test (FET) with the zebrafish (*Danio rerio*) a potential alternative for the fish acute toxicity test? *Comp Biochem Physiol C: Pharmacol Toxicol Endocrinol*, 149, 196-209, (2009).
- (28) T. LUCKENBACH, M. KILIAN, R. TRIEBSKORN, O. OBEREMM, Fish early life stage tests as a tool to assess embryotoxic potentials in small streams. *J Aquat Ecosyst Stress Recovery*, 8, 355–370, (2001).
- (29) DIRECTIVE 2010/63/EU of the European Parliament and of the Council of the European Union. On the Protection of Animals Used for Scientific Purposes. *OJEU*. L276, 233–279, (2010).
- (30) TSO (The Stationary Office), Guidance on the Operation of the Animals (Scientific Procedures) Act, London, (2000).
- (31) J.A. CAMARGO, A. ALONSO, Ecological and toxicological effects of inorganic nitrogen pollution in aquatic ecosystems: A global assessment. *Environ Int*, 32, 831-849 (2006).
- (32) E. SCHRAM, J.A.C. ROQUES, T. VAN KUIJK, W. ABBINK, J. VAN DE HEUL, P. DE VRIES, S. BIERMAN, H. VAN DE VIS, G. FLIK, The impact of elevated water ammonia and nitrate concentrations on physiology, growth and feed intake of pikeperch (*Sander lucioperca*), *Aquaculture*, 420–421, 95-104 (2014).
- (33) S. WUERTZ, S.G.E. SCHULZ, U. EBERHARDT, C. SCHULZ, J.P. SCHROEDER, Acute and chronic nitrite toxicity in juvenile pike-perch (*Sander lucioperca*) and its compensation by chloride, *Comp Biochem Physiol*, 157, 352-360, (2013).
- (34) L. NOEL, R. CHEKRI, S. MILLOUR, M. MERLO, J.C. LEBLANC, T. GUERIN, Distribution and relationships of As, Cd, Pb and Hg in freshwater fish from five French fishing areas, *Chemosphere*, 90, 1900-1910 (2013).
- (35) S. SUBOTIC, S. SPASIC, Z. VISNJIC-JEFTIC, A. HEGEDIS, J. KRPO-CETKOVIC, B. MICKOVIC, S. SKORIC, M. LENHARDT, Heavy metal and trace element bioaccumulation in target tissues of four edible fish species from the Danube River (Serbia). *Ecotox Environ Saf*, 98,196-202, (2013).
- (36) WASZAK, H. DABROWSKA, Persistent organic pollutants in two fish species of *Percidae* and sediment from the Sulejowski Reservoir in central Poland, *Chemosphere*, 75, 1135-1143, (2009).